

PRENYL SUBSTITUTED CYCLOBUTANONES AS SQUALENE
SYNTHETASE INHIBITORS¹

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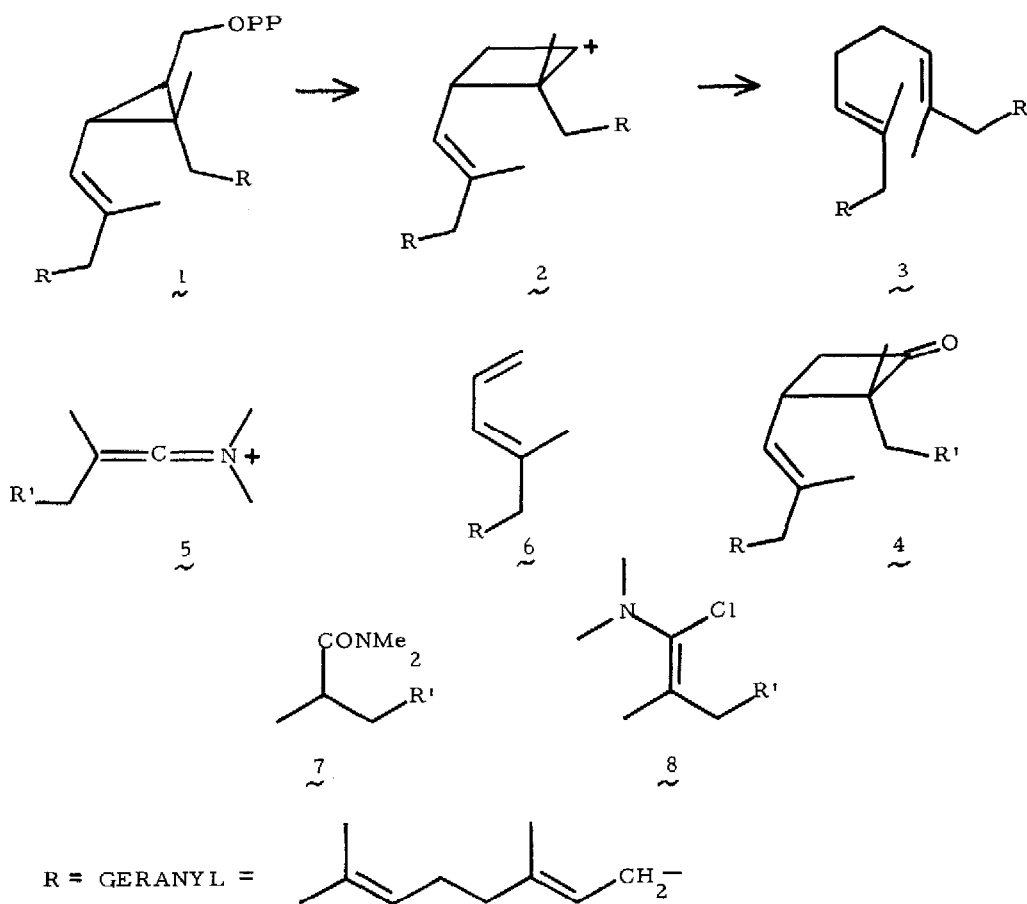
Although virtually no direct information is available on the enzymatic mechanism by which presqualene pyrophosphate (1) is transformed into squalene (3),² solvolysis studies with small (ten carbon) model compounds have been used to define thermodynamically favored pathways.³ These formally entail pyrophosphate loss to a cyclopropylmethylene carbonium ion, multiple rearrangement to the squalene skeleton, and quenching of the cation by hydride transfer. The model studies suggest that the highest energy barrier in the rearrangement sequence lies near cyclobutyl cation 2,^{3a} the pyrophosphate of which has been mentioned as a possible discrete intermediate.⁴ If model studies do, in fact, reflect the enzymatic mechanism, the transition state for the enzymatic rearrangement sequence should also approximate 2.⁵ Cyclobutanone analogs of cation 2, in which the charged atom is replaced by a similarly hybridized and polarized carbonyl carbon, might therefore function as "transition state" inhibitors of the enzyme squalene synthetase.⁶ The cyclobutanones would also be useful as precursors to cyclobutanol derivatives for mechanistic and model studies. We report here the synthesis and preliminary bioassay of 3-(2,6,10-trimethylundeca-1,5,9-trienyl)-2,2-dimethylcyclobutanone (4a) and 3-(2,6,10-trimethylundeca-1,5,9-trienyl)-2-methyl-2-(4,8-dimethyl-1-nonyl)cyclobutanone (4b), as well as a functional group incompatibility in 5c which has prevented synthesis of 4c.

The cyclobutanones were assembled by polar cycloaddition of keteneimmonium cations⁷ 5a and 5b to 4,8,12-trimethyl-1,3,7,11-tridecatetraene (6). Diene 6 was prepared in 94% yield by Wittig condensation of methylenetriphenylphosphorane and E,E-farnesal.⁸ N,N-Dimethyl-1-chloro-2-methylpropenylamine (8a), the precursor of 5a, was prepared from N,N-dimethylisobutyramide (7a)⁹ by sequential treatment with gaseous phosgene¹⁰ and triethylamine.⁷ Addition of 8a

(3 mmol) to a CH_2Cl_2 solution of **6** (2.99 mmol) and AgBF_4 (3.23 mmol) at -78°C , warming the mixture slowly to room temperature, filtration, aqueous base workup, silica gel chromatography, and bulb-to-bulb distillation gave cyclobutanone **4a** (59%), a single peak on glc (13.99 min, 175°C);¹¹ ir (film) 1790 cm^{-1} ; nmr (CDCl_3) 1.03 and 1.22 (s, 6H, cyclobutyl methyls), 1.63 and 1.68 (s, 12H, allyl methyls), 1.88–2.23 (m, 8H, allyl methylenes), 2.68–3.35 (m, 3H, cyclobutyl H), and 4.87–5.33 ppm (m, 3H, vinyl H); CIMS m/e 289 (molec. ion + 1). Anal. ($\text{C}_{20}\text{H}_{32}\text{O}$): calcd., C, 83.27; H, 11.18; found C, 82.98, H, 11.05. The nmr signal at 1.03 ppm is attributed to the cyclobutyl methyl cis to the farnesyl group, and that at 1.21 ppm to the trans methyl, by correlation with pertinent literature examples.¹²

The α -chloroamine **8b** was prepared from dimethylamide **7b** in 99% yield (allowing for recovered starting material) as described for **8a**.⁸ Amide **7b** was obtained by catalytic hydrogenation of **7c** (10% Pd/C, ethanol, 97.5% yield), which in turn was made in 92% yield by stepwise reaction of the known free acid¹³ with 1,1-carbonyldiimidazole and dimethylamine.^{8,14} Treatment of **8b** and **6** with AgBF_4 in CH_2Cl_2 , as described above, gave a mixture of products from which **4b** (cyclobutyl Z:E ratio 6:4 by glc) was isolated in 35% yield by high vacuum removal of volatile components and silica gel chromatography: ir (film) 1785 cm^{-1} ; CIMS m/e 429 (molec. ion + 1); calc. exact mass for $\text{C}_{30}\text{H}_{52}\text{O}$, 428.4018; observed 428.4016.¹⁵ The isomer mixture, although difficult to separate, gave each isomer with 30% contamination of the other on careful column chromatography through silica gel. These enriched mixtures showed that the two isomers had similar nmr spectra except for the position of the cyclobutyl methyl signals. The common portion of the spectrum is as follows: 0.87 (d, $J=5.8\text{ Hz}$, 9H, aliphatic methyls), 0.96–1.54 (m, 14H, aliphatic CH_2 and CH), 1.62 and 1.68 (s, 12H, allylic methyls), 1.70–2.40 (m, 8H, allylic CH_2), 2.60–3.34 (m, 3H, cyclobutyl H), and 5.17 ppm (m, 3H, vinyl H). The cyclobutyl methyl group appeared at 1.21 ppm in the major isomer of the product mixture (glc 8.70 min, 240°C),¹¹ and at 1.03 ppm in the minor isomer (glc 10.01 min, 240°C).¹¹ Correlation of these signals with those of the methyl groups in **4a** allows assignment of the cyclobutyl-Z configuration to the major isomer.

Synthesis of **4c** by analogous condensation of **3c** with **6** gave a complex mixture, in which products with infrared absorption appropriate to cyclobutanones did not have thirty-carbon skeletons. Analysis of the product mixture by nmr indicated substantial loss of vinyl protons, suggesting that intramolecular cyclizations of **5c** may have occurred. The use of keteneimmonium cations for cyclobutanone synthesis thus appears limited to compounds without vicinal unsaturation.



R' = a) H b) TETRAHYDROGERANYL c) GERANYL

Cyclobutanones 4a and 4b are weak inhibitors of yeast squalene synthetase compared with farnesyl pyrophosphate analogs.¹⁶ A 10-15% inhibition was observed in our bioassay¹⁶ with the cyclobutanones at about 5 mM concentration, although the quantitative accuracy of the data is clouded by the water insolubility of the compounds.

REFERENCES AND NOTES

- (1) a) This work was supported by grant HL-15476 from the National Institutes of Health b) We thank Dr. Jeng Shu Wei for the bioassay data.
- (2) a) E. Beytia, A. A. Qureshi, and J. W. Porter, *J. Biol. Chem.*, 248, 1856 (1973). b) P. R. Ortiz de Montellano, R. Castillo, W. Vinson, and J. S. Wei, *J. Am. Chem. Soc.*, 98, 3020

(1976).

- (3) For excellent reviews of the model studies see a) C. D. Poulter, J. Agr. Food Chem., 22, 167 (1974). b) C. D. Poulter, O. J. Muscio, and R. J. Goodfellow, Biochem., 13, 1530 (1974).
- (4) J. Edmond, G. Popjak, S. M. Wong, and V. P. Williams, J. Biol. Chem., 246, 6254 (1971).
- (5) G. S. Hammond, J. Am. Chem. Soc., 77, 334 (1955).
- (6) a) G. E. Lienhard, Science, 180, 149 (1973). b) R. Wolfenden, Accts. Chem. Res., 5, 10 (1972). c) R. N. Lindquist, Chapter 2 in "Drug Design", Vol. 5, E. J. Ariens, Ed., Acad. Press, N. Y., 1975.
- (7) J. Marchand-Brynaert and L. Ghosez, J. Am. Chem. Soc., 94, 2870 (1972).
- (8) The ir, nmr, and chemical ionization mass spectrum of all isolated compounds were in complete agreement with the assigned structures. All new compounds, except 6, which air oxidized readily, gave satisfactory elemental analyses.
- (9) a) H. Rapoport and R. M. Bonner, J. Am. Chem. Soc., 72, 2783 (1950) b) Y. Miron and H. Morawetz, Macromolec., 2, 162 (1969).
- (10) Use of commercial phosgene-benzene solutions instead of phosgene gas severely diminished yields.
- (11) GLC on a Varian 2100 with flame ionization detectors and 6 ft x 2 mm i.d. 3% OV-225 on 100-200 mesh Varoport 30 all-glass columns, at a nitrogen flow of 18 ml/min.
- (12) a) R. M. Coates and W. H. Robinson, J. Am. Chem. Soc., 94, 5920 (1972) b) J. Grandguillot and F. Rouessac, Compt. Rend. Acad. Sci., Ser. C, 277, 1253 (1973).
- (13) K. Mori and M. Matsui, Tetrahedron, 26, 2801 (1970).
- (14) H. A. Staab and K. Wendel, Org. Synth., 48, 44 (1968).
- (15) We thank Prof. A. Burlingame, Space Sciences Lab., Berkeley, for exact mass determinations on an AEI-MS902.
- (16) P. R. Ortiz de Montellano, J. S. Wei, R. Castillo, C. K. Hsu, and A. Boparai, J. Med. Chem., in press.